
Medicine-Based Evidence: Reverse Translational Ear Research Recommendations

Kenneth H. Brookler

Lenox Hill Hospital, New York, New York, USA

Abstract: Presented here is a first-person account of the evolution of the practice of surgical neurotology to that of medical neurotology shaped mainly by results of treatment directed at underlying otosclerosis-like lesions of the otic capsule and metabolic factors. With new technologies and rapidly evolving concepts, the changing treatment algorithms did not remain constant to provide the usual evidence-based outcome analyses. However, the majority of the patients presenting with neurotological symptoms had undergone previous medical or surgical treatment before undergoing the medical management herein described. The underlying ongoing basic science findings over this period were linked to the clinical observations. On the basis of the more effective results of treating neurotological disorders, recommendations are made for future areas of investigation—mostly basic science—into developing an investigative foundation for future effective management of patients with a variety of neurotological disorders.

Key Words: hyperinsulinemia; impaired glucose tolerance; Ménière's syndrome; osteoclast; osteoprotegerin; otosclerosis-like

For the last 40 years, I have been immersed in the field of clinical neurotology. My roots were in two distinct disciplines. The first discipline is the study of the underlying mechanisms of clinical problems in neurotology. The second discipline is surgical neurotology.

After finishing a surgically oriented neurotology fellowship in 1968, my next pursuit was to find surgical cases, almost to the exclusion of any nonsurgical cases. I was particularly focused on the surgical treatment of dizziness and Ménière's syndrome [1–4]. At that time, no treatment for progressive sensorineural hearing loss or tinnitus was available. As my clinical surgical experience grew, I was fortunate to be influenced by others to return to my roots and explore the underlying mechanisms of clinical disease. My treatments began to change in response to my continuing observations and to new developments in basic science and clinical practice.

Two main treatment pathways emerged as a basis for my medical (nonsurgical) treatment of neurotological disorders [5–8]. One treatment pathway was based on

the recognition of an inner-ear otosclerosis-like disorder that was discovered while searching for an explanation for an etiology underlying the clinical findings. The other treatment pathway focused on the metabolism of the inner ear. In time, my experience with these treatment algorithms made me realize that some medical solutions are more effective than surgical solutions. In fact, the medical managements that evolved were disease-modifying, far more efficient, more cost-effective, and less morbid than the invasive surgical alternatives and previous and current medical management standards of practice.

PATIENTS, METHODS, AND RESULTS

The decisive factor in achieving successful outcomes in patients with neurotological symptoms is the manner in which hair cell function is modified throughout the duration of treatment. Normal hair cell function is dependent on the muscle or actin component of the hair cell. Current hair cell research is focused on the motor protein prestin (which is found within the actin), on repopulating or replacing hair cells, and on actin elements in general.

Prestin research has given us a greater understanding of the cochlear amplifier of the outer hair cells [9–13]

Reprint requests: Kenneth H. Brookler, MD, Lenox Hill Hospital, 111 East 77th Street, New York, NY 10075. Phone: 212-861-6901; Fax: 212-472-0134; E-mail: khbrooklermd@mac.com

and the underlying movement of calcium in the changed polarization of the cochlear hair cells [14,15] at the synaptic junction. Repopulation has focused on the conversion of supporting cells or on the unlocking of the genetic code observed in lower species that allows for repopulation. Hair cell replacement is investigated in stem cell research.

Impaired function of hair cells is attributable to a disorder in the behavior of the actin elements. When hair cell function is impaired, as demonstrated clinically by otoacoustic emissions testing, the dysfunction appears to be the source of the otological and neurootological symptoms we treat in practice. If hair cell function can be improved, the symptoms that go along with impaired function can abate or disappear. In some of my patients, hearing loss has been significantly reversed. With the inclusion of routine otoacoustic emission testing in clinical practice, reversible otoacoustic emission results infer regeneration of hair cell function along with parallel clinical improvement. Though the thrust in current hair cell research is the repopulation of hair cells or their replacement with stem cells, little research has been directed at recovery of impaired hair cell function.

Analysis of clinical observations along with the results of evolving basic research could explain the clinical findings in patients with neurootological symptoms and lead to novel clinical treatments. Thus, this article serves as my proposal for future ear research.

Clinical Treatment Observations

The results of my medical treatments led me to develop further innovations, which in turn prompted more questions. Some of the answers to these questions were found in the domain of basic science. I have been awed, at meetings of the Association for Research in Otolaryngology, by the presentations of basic science methodologies used to investigate the processes of inner-ear function. However, as basic science techniques continue to progress, it is important that we not overlook some of the older investigative techniques that could help explain the underlying mechanisms of some of my treatment successes. Though it is true that some of the newer investigative techniques have validated some of my observations, more research using both older and newer techniques is required to further advance the successful management of these patients presenting with neurootological symptoms.

Clinical Observations of Otosclerosis

As mentioned, my first treatment pathway was based on my understanding that underlying otosclerosis or otosclerosis-like lesions of the inner ear can produce a

variety of symptoms, including Ménière's syndrome, dizziness, sensorineural hearing loss, tinnitus, hyperacusis, and some hemicranial headaches [16]. When I was in training, those who studied the influence of otosclerosis on the inner ear were divided into two camps. One camp believed that the otosclerotic lesion must impinge on the endosteal membrane [17,18] to alter inner-ear function [19,20], whereas the other camp believed that the mere presence of otosclerosis in the temporal bone could influence the inner ear [21–25]. By 2005, we found the existence of lacunar and canalicular channels that lead from the perilymphatic space into the otic capsule [26]. The presence of these channels explains how the by-products of the otosclerotic process can enter the perilymph to exert their potential effect on the sensory structures of the inner ear without the otosclerotic focus physically impinging on the endosteal membrane. By 2008, we found that temporal bone histopathology explained some of the variety of clinical findings [27–30].

When I finished my fellowship in 1969, sodium fluoride was the only medical otosclerosis treatment that was recognized by some authors [31–40]. However, its use was controversial at that time because of general nutritional considerations and a divided clinical community [41,42]. During my training, I was not firmly convinced of the value of sodium fluoride in the treatment of otosclerosis. However, my fellowship lasted only 1 year, and this may not have been enough time for me to arrive at any conclusion about the efficacy of sodium fluoride in the medical treatment of the neurootological symptoms of otosclerosis.

In my search for candidates for acoustic tumor surgery in the early 1970s, I obtained tomographic scans of the temporal bones to more accurately examine the size of the internal auditory canals. Though most of these acoustic neuroma candidates did not demonstrate findings that would suggest an acoustic tumor, many of them had polytomographic evidence of otosclerosis [43,44]. Those who did demonstrate polytomographic evidence of otosclerosis were placed on sodium fluoride to treat the neurootological symptoms. As a result of the sodium fluoride treatment, many patients showed evidence that their Ménière's syndrome had stabilized or that their vertigo or progressive sensorineural hearing loss had been arrested. Some patients also demonstrated a reduction in their tinnitus and hearing loss [45].

On the basis of these observations, I began treating my otosclerosis patients with etidronate after the bisphosphonate group of drugs became available (around 1990) [16]. Etidronate was approved for the treatment of Paget's disease of the bone and demonstrated its effect on the hearing loss in Paget's disease [46]. Etidronate was subsequently studied as a treatment for osteoporosis. It seemed logical that this compound could also be

considered for the treatment of otosclerosis. Indeed, etidronate quickly demonstrated that it was more effective than sodium fluoride alone for the treatment of otosclerosis. My success with etidronate led me to use other bisphosphonates to treat the inner-ear symptoms of otosclerosis [16]. Newer generations of bisphosphonates became available, and their effectiveness became predictable. They were better than sodium fluoride alone and better than any surgical procedure directed at the inner-ear symptoms of otosclerosis.

Basic Science of Otosclerosis

Our understanding of the active phase of otosclerosis is that it involves an osteoclast-driven dystrophy of the otic capsule bone. In the basic science world, the field of osteoclastogenesis developed around the turn of the twenty-first century. The inner ear was found to produce a cytokine called *osteoprotegerin* (OPG) [26]. OPG acts as a decoy for the receptor activator of nuclear kappa B ligand, another cytokine that attaches at its receptor site to a macrophage, transforming it into an osteoclast. In addition, channels that were identified connect the perilymphatic space with the otic capsule, a finding that could explain how OPG diffuses into the otic capsule [26] and how the byproducts of the breakdown of bone (osteoclast activity) could diffuse in a direction opposite from that of the otic capsule and into the perilymph to produce inner-ear symptoms. Without a membrane barrier, these channels provide a conduit for OPG to diffuse out into the otic capsule bone and prevent or reduce osteoclast production. This effect results in very low bone turnover, which makes the normal otic capsule the hardest bone in the body [47]. In the past, some believed that the otosclerotic area had to impinge on the endosteal membrane for neurootological symptoms to occur [42]. The cytokines that are breakdown products of bone enter into the perilymph through the same channels that the OPG uses to diffuse out into the otic capsule.

Tumor necrosis factor alpha (TNF- α) is one of the cytokine byproducts of osteoclastic activity that has been shown to be toxic to the inner ear, presumably to the hair cells [48–57]. An investigation of mice that were genetically altered so as not to produce OPG (OPG knockout mice) found that hearing was lost shortly after birth; histopathology of the temporal bones revealed lesions similar to those seen in humans with otosclerosis [58]. A recent investigation revealed that intraperitoneal bisphosphonate (risedronate) in OPG knockout mice prevented the development of otosclerosis-like lesions and hearing loss [59]. Because bisphosphonates target osteoclasts and because otosclerosis or otosclerosis-like disorders are osteoclast-driven, the use of bisphosphonates could truly produce a disease-modifying effect.

For the last 3 years, my laboratory evaluations have included the identification of 25-hydroxy vitamin D. I look at vitamin D₃ and parathyroid hormone (PTH) levels in particular. This has allowed me to identify deficient (<20 ng/ml) and insufficient (<40 ng/ml) vitamin D levels and primary and secondary hyperparathyroidism even in patients whose serum calcium levels are normal. In some patients, hyperparathyroidism occurred secondary to insufficient vitamin D and was reversible by appropriate vitamin D replacement [6,60–62]. Vitamin D and PTH abnormalities may have an effect on osteoclastogenesis, on the neurootological symptoms of otosclerosis [60–62], and on the efficacy of bisphosphonates. Patients who present with a radiological finding of a superior semicircular canal fistula (with or without symptoms) have been found to have abnormalities of vitamin D or PTH levels or both. When these abnormalities are identified and treated, many patients experience a regression of symptoms or an improvement of otoacoustic emissions. My management of superior semicircular canal dehiscence suggests that it may represent a metabolic disorder related to insufficient vitamin D or to primary or secondary hyperparathyroidism. The fact that it may not be a surgical disorder may explain why it can recur in patients who have undergone surgery to replace the bone. It would also explain why canal plugging is a more effective surgical treatment. In my experience, some patients who undergo surgery for chronic otitis media with cholesteatoma have been found to have an eroded and exposed dehiscent lateral semicircular canal. The cholesteatoma matrix is peeled off the lateral semicircular canal fistula and, after an interval when a second operation for hearing restoration has been performed, the bone has usually regrown over the lateral semicircular canal. If superior semicircular canal dehiscence is a metabolic disorder of bone, the bony dehiscence may in fact spontaneously close, given the correct conditions of vitamin D or PTH (or both).

Moreover, carbonic anhydrase, an enzyme in the stria vascularis [63–77], has been speculated to play a role in electrolyte homeostasis in the inner ear. Acetazolamide is a carbonic anhydrase inhibitor that is used as a diuretic; in fact, it is frequently the diuretic of choice for treating otological symptoms [78–81]. Carbonic anhydrase also plays a role in osteoclastogenesis. Osteoclasts express an mRNA for carbonic anhydrase [82–89]. Carbonic anhydrase is found at the ruffled border of the osteoclast. Presumably, its function is to provide the hydrogen ion required to produce the acid needed to break down bone. PTH stimulates carbonic anhydrase activity, thereby producing the acidity necessary for osteoclastic activity. It is possible that carbonic anhydrase production may play a reciprocal role with the production of OPG by the inner ear.

The molecular biology of bone in general and otosclerosis-like lesions in the experimental animal and in humans explains how patients may demonstrate radiographic evidence of otosclerosis and respond to bisphosphonate treatment without experiencing the typical symptoms associated with otosclerosis.

Clinical Observations of Metabolic Aspects of Inner-Ear Disorders

The evolution of my second treatment pathway had to do with the energy requirements of the inner ear. During the late 1960s and mid-1970s, information was developing about an effect that blood glucose and lipids had on the inner ear [90–104]. Other lines of thought included concepts of thyroid disease [105,106] and evidence of insulin disorders that could influence blood glucose levels and their effect on inner-ear function. In 1992, published criteria for considering abnormally elevated insulin levels on a glucose tolerance test were developed [102]. Patients presenting with clinical neurootological symptoms were evaluated for thyroid disease, lipid disorders, and blood glucose and insulin disorders with the use of a 5-hour glucose tolerance test with simultaneous insulin levels [107]. The most common abnormalities were seen on the glucose tolerance test. Though only a few patients exhibited evidence of diabetes or hypoglycemia, many of the remainder showed abnormal fluctuations in blood glucose levels; in rare cases, a flat curve in the blood glucose level was seen [103]. These glucose levels were sufficiently abnormal to adversely affect the function of an already impaired inner ear, but they did have a high probability of responding favorably to dietary management.

Coincident with these blood glucose findings were findings of hyperinsulinemia [108–110]. During this time, endocrinologists identified the insulin resistance syndrome, which was first called *syndrome X* and, later, *metabolic syndrome* [111]. By virtue of the highly metabolic nature of the inner ear, especially the hair cells, subtle changes in blood glucose or elevations in insulin could explain the genesis of some inner-ear symptoms. In addition, in patients so identified, the adversely functioning inner ear acts as the “canary in the coal mine” by identifying patients who are at risk of developing insulin resistance syndrome and its sequelae. Untreated, the insulin resistance syndrome can progress to such diseases as non-insulin-dependent (type 2) diabetes [112,113], hypertension [104], cardiovascular disease, and non-alcoholic fatty liver disease.

Recent literature has suggested a possible convergence of my two treatment pathways. Research in bone metabolism and regulation has demonstrated an overlap with insulin regulation. One has to do with a plasma cell membrane glycoprotein-1 (PC-1) [114,115] that is

involved in bone metabolism and may also be involved in regulating the entry of insulin into cells [116–127]. Other findings suggest that bone in general and osteocalcin in particular may be potent insulin regulators [128–136].

Research has also demonstrated that insulin and blood glucose have an effect on inflammatory markers and OPG and a possible effect on otic capsule bone. Fetuin-A is a hepatic secretory protein that binds to the insulin receptor and inhibits insulin action. High levels are associated with insulin resistance and with incident diabetes independent of physical activity, inflammatory markers, and other common markers of insulin resistance [137–140]. Fetuin-A may play a role in hair cell function, and it is known to be an inhibitor of extraosseous calcification. In addition, a relationship between fetuin-A and OPG has been described [141,142]. These insulin and osseous effects may bring together a concept of the convergence of carbohydrates in the diet and their effect on blood glucose, insulin, and osteoclast function as evidenced by otosclerosis-like lesions. Peroxisome proliferator-activated receptor-gamma (PPAR- γ) agonist belongs to the nuclear receptor superfamily composed of ligand-activated transcription factors. It is highly expressed in adipose tissue and involved in insulin sensitization. It has also been shown to inhibit many inflammatory mediators, including TNF- α and interleukin-6. It has been shown to be an important regulator of airway inflammation and allergic rhinitis. Rosiglitazone is a PPAR- γ agonist used primarily to decrease insulin resistance in type 2 diabetes. Some animal experiments showed that it has a neuroprotective effect in experimental stroke [143]. Because of this effect on inflammatory mediators, rosiglitazone may inhibit the triggers of osteoclast production and could be considered as a replacement for corticosteroids in the treatment of autoimmune disorders of the inner ear.

Molecular Biology of Otosclerosis and Effects of Inflammatory Cytokines on the Inner Ear

Specimens obtained from patients who had previously undergone stapedectomy to treat the conductive hearing loss of otosclerosis showed that they clearly had otosclerosis. These specimens from stapedectomy procedures have shown the presence of TNF- α in addition to strong evidence of prior measles virus exposure [52]. Another study in experimental animals demonstrated that the use of a TNF- α -neutralizing agent could prevent the sensorineural hearing loss secondary to induced pneumococcal meningitis [48]. Other authors have reported the role of inflammatory cytokines, particularly TNF- α , in inner-ear inflammation [49,54,55]. The field of osteoclastogenesis informs us that one of the byproducts of osteoclast

production and the breakdown of bone is TNF- α [50, 57,144–146].

Vestibulocollic Headache and Vestibular “Migraine”

I became interested in a reflex that, in a number of patients, was initiated by the vestibular portion of the inner ear and resulted in muscular contraction primarily focused in the cervical area. Many of these patients had evidence of spasm in the muscles of the neck behind the mastoid, usually on the same side as the vestibular abnormality. Some of these patients also had headache, usually hemicranial. Some patients had only hemicranial headache and no vestibular symptoms, yet they showed signs of abnormal vestibular function [147].

Review of the pertinent literature related to migraine reveals that its fundamental mechanism is still not fully understood. It is possible that in a large number of migraineurs, migraine may be a manifestation of an abnormally functioning vestibular system that does not necessarily cause vestibular symptoms. It is only by evaluating the vestibular system in these headache patients that an abnormality could be identified.

RECOMMENDATIONS FOR FUTURE RESEARCH

I make the following recommendations for research, which are based on my many years of clinical observations and my review of the basic science and other clinical literature relevant to these findings.

Otosclerosis

We should continue to investigate the effects of the bisphosphonates on OPG knockout mice. If we can identify a genetic line of experimental animal that develops progressive sensorineural hearing loss, we can measure the levels of OPG over time to determine whether the inner ear produces less OPG and whether this could be a factor in progressive sensorineural hearing loss that, in the past, was attributed to aging or another process.

We should also experiment with mouse monoclonal antibody to OPG. By selectively blocking or reducing the production of OPG, this antibody could be used experimentally to determine whether hearing loss or histopathological evidence of otosclerosis-like lesions can be induced by progressive alterations in this antibody.

We need to develop imaging guidelines for radiologists to use in the early diagnosis of otosclerosis-like lesions. In addition to measuring otoacoustic emissions in OPG knockout mice and performing postmortem histopathology of hair cells (including mitochondria and pres-

tin), clinicians should obtain computed tomographic scans of the temporal bones. Studies of OPG knockout mice have revealed that they have histopathological lesions similar to those seen in human otosclerosis. This finding suggests that otosclerosis-like lesions may be triggered in humans by something other than the measles virus [51,58,148,149].

Practitioners and researchers should investigate the effect of PTH on the otic capsule to determine whether it can be used as a treatment for otic capsule disorders or plays any role in otic capsule disorders. Likewise, otoconia should be investigated in relation to OPG, vitamin D, and PTH. Investigations into vitamin D deficiency or insufficiency may help us determine whether this vitamin has any effect on the otic capsule in terms of osteoclastogenesis, inner-ear OPG, or hyperparathyroidism.

By inducing a vitamin D deficiency or hyperparathyroid state in experimental animals, we may be able to determine whether superior semicircular canal dehiscence will develop. If so, we can investigate whether the dehiscence will close once vitamin D or PTH levels (or both) are restored to normal.

Finally, we should investigate the effect of acetazolamide alone, with PTH, and with bisphosphonates on otoacoustic emissions in wild mice and then in OPG knockout mice.

METABOLIC ABNORMALITIES OF THE INNER EAR

The viscosity of endolymph and perilymph warrants investigation. Knowledge of the viscosity can lead to further understanding of the tuning of the inner ear to sound. In addition, it is likely that the viscosity of the endolymph is such that otoconia may not freely move in endolymph with a change in the position of the head, which would contradict the current explanation for benign paroxysmal positional vertigo and nystagmus.

The production of endolymph also warrants study. Is there a feedback mechanism between the hair cells (particularly with regard to prestin) and the secretory mechanism of endolymph that creates the proper pressure medium for the most efficient hair cell function? If inefficient functioning reduces the rigidity of hair cells, is there a reflex stimulus to secrete more endolymph to overcome the necessary pressure? Would this explain the increase in the endolymphatic volume in Ménière’s syndrome without an alteration in the sodium and potassium components of the endolymph?

Investigators should produce hypoglycemia in experimental animals and test for otoacoustic emissions, for changes in the endocochlear potential, and for postmortem changes in the mitochondria of the hair cells and supporting cells. They might also evaluate and compare

otoacoustic emissions and mitochondria size in the hair cells of Zucker rats and wild rats. Zucker rats produce type 2 diabetes, and they have been found to undergo changes in the mitochondria of skeletal muscle. Possibly the effects of diabetes on the inner ear may, in turn, have an effect on the mitochondria of the hair cells.

Researchers should induce blood glucose abnormalities in OPG knockout mice and measure any additional effect they may have on mitochondria and prestin function. A baseline investigation of the mitochondria before the blood glucose abnormalities are induced could also show a possible effect of TNF- α on the hair cells. Research projects should be integrated to include investigations of PC-1, fetuin-A, and osteocalcin, as there is a convergence of both metabolic and osteoclast functions. In turn, these findings should be integrated with what we know about mitochondria, hair cell actin, endocochlear potential, endolymph viscosity, and actin (prestin) of hair cells and the role of TNF- α in inner-ear function.

SUMMARY

It is my enthusiastic hope that the information contained herein will spark a team or teams of scientists to further investigate the observations as yet unexplained. Finding expertise in basic science investigation in one laboratory appears unlikely. Regenerating hair cell function through the use of clinical management algorithms built on a basic research foundation could lead to the development of effective treatments for a variety of inner-ear disorders. Effective treatment of inner-ear disorders will likely stimulate more questions that can be answered only through basic science investigations.

ACKNOWLEDGMENT

The author thanks Martin Stevenson for his help in preparing the manuscript.

REFERENCES

- Hoffman RA, Brookler KH, Reich EJ. Trigeminal neuralgia symptomatic of acoustic neuroma. *N Y State J Med* 79:1436–1438, 1979.
- Miyamoto RT, Althaus SR, Wilson DF, Brookler KH. Middle fossa surgery. Report of 153 cases. *Otolaryngol Head Neck Surg* 93:529–535, 1985.
- Brookler KH, Hoffman RA, Camins M, Terzakis J. Tri-lobed meningioma: Ampulla of posterior semicircular canal, internal auditory canal, and cerebellopontine angle. *Am J Otol* 1:171–173, 1980.
- Brookler KH. Acoustic neuroma surgery. *Am J Otol* 3: 46–50, 1981.
- Brookler KH. Ménière's disease. Role of otospongiosis and metabolic disorders. *Acta Otolaryngol Suppl* 406: 31–36, 1984.
- Brookler KH, Glenn MB. Ménière's syndrome: An approach to therapy. *Ear Nose Throat J* 74:534–538, 540, 542, 1995.
- Brookler KH, Rubin W. The dizzy patient: Etiologic treatment. *Otolaryngol Head Neck Surg* 103:677–680, 1990.
- Rubin W, Brookler KH. Etiologic diagnosis in neurotologic disease. *Otolaryngol Head Neck Surg* 103:693–694, 1990.
- Zheng J, Shen W, He DZ, et al. Prestin is the motor protein of cochlear outer hair cells. *Nature* 405:149–155, 2000.
- Zheng J, Madison LD, Oliver D, et al. Prestin, the motor protein of outer hair cells. *Audiol Neurootol* 7:9–12, 2002.
- Dallos P. Cochlear amplification, outer hair cells and prestin. *Curr Opin Neurobiol* 18:370–376, 2008.
- Santos-Sacchi J, Navarrete E, Song L. Fast electro-mechanical amplification in the lateral membrane of the outer hair cell. *Biophys J* 96:739–747, 2009.
- Yu N, Zhu ML, Zhao HB. Prestin is expressed on the whole outer hair cell basolateral surface. *Brain Res* 1095: 51–58, 2006.
- Canis M, Schmid J, Olzowy B, et al. The influence of cholesterol on the motility of cochlear outer hair cells and the motor protein prestin. *Acta Otolaryngol* Jan 19: 1–6, 2009.
- Frolenkov GI. Ion imaging in the cochlear hair cells. *Methods Mol Biol* 493:381–399, 2009.
- Brookler KH, Tanyeri H. Etidronate for the the neurotologic symptoms of otosclerosis: Preliminary study. *Ear Nose Throat J* 76:371–376, 379–381, 1997.
- Schuknecht HF, Kirchner JC. Cochlear otosclerosis: Fact or fantasy. *Laryngoscope* 84:766–782, 1974.
- Schuknecht HF. Myths in neurotology. *Am J Otol* 13: 124–126, 1992.
- Schuknecht HF. Cochlear otosclerosis. A continuing fantasy. *Arch Otorhinolaryngol* 222:79–84, 1979.
- Schuknecht HF. Cochlear otosclerosis. An intractable absurdity. *J Laryngol Otol Suppl* 8:81–83, 1983.
- Linthicum FH Jr, Neely JG. Unrelated sensorineural hearing loss in patients with otosclerosis. A report of three cases. *Laryngoscope* 87:1746–1752, 1977.
- Linthicum FH Jr, Lalani AS. Sensorineural impairment in unilateral otosclerosis. *Ann Otol Rhinol Laryngol* 84: 11–15, 1975.
- Linthicum FH, Ear Research Institute. *Cochlear Otosclerosis Histology and Physiology*. Los Angeles: The Institute, 1975.
- Linthicum FH Jr. Diagnosis of cochlear otosclerosis. *Arch Otolaryngol* 95:564–569, 1972.
- Kelemen G, Linthicum FH Jr. Labyrinthine otosclerosis. *Acta Otolaryngol Suppl* 253:1–68, 1969.
- Zehnder AF, Kristiansen AG, Adams JC, et al. Osteoprotegerin in the inner ear may inhibit bone remodeling in the otic capsule. *Laryngoscope* 115:172–177, 2005.

27. Makarem A, Linthicum FH. Cochlear otosclerosis and endolymphatic hydrops. *Otol Neurotol* 29:571–572, 2008.
28. Makarem AO, Linthicum FH. Cavitating otosclerosis. *Otol Neurotol* 29(5):730–731, 2008.
29. Balle V, Linthicum FH Jr. Histologically proven cochlear otosclerosis with pure sensorineural hearing loss. *Ann Otol Rhinol Laryngol* 93:105–111, 1984.
30. Abd el-Rahman AG. Cochlear otosclerosis: Statistical analysis of relationship of spiral ligament hyalinization to hearing loss. *J Laryngol Otol* 104:952–955, 1990.
31. Linthicum FH Jr, Forquer BD. Sodium fluoride as a treatment for otosclerotic hearing loss. *Am J Otol* 6:35–37, 1985.
32. Linthicum FH Jr, House HP, Althaus SR. The effect of sodium fluoride on otosclerotic activity as determined by strontium. *Ann Otol Rhinol Laryngol* 82:609–615, 1973.
33. Shambaugh GE Jr. How and when to prescribe sodium fluoride. *Am J Otol* 10:146–147, 1989.
34. Forquer BD, Linthicum FH, Bennett C. Sodium fluoride: Effectiveness of treatment for cochlear otosclerosis. *Am J Otol* 7:121–125, 1986.
35. Causse JR, Causse JB, Uriel J, et al. Sodium fluoride therapy. *Am J Otol* 14:482–490, 1993.
36. Causse JR, Shambaugh GE, Causse B, Bretlau P. Enzymology of otospongiosis and NaF therapy. *Am J Otol* 1:206–214, 1980.
37. Bretlau P, Salomon G, Johnsen NJ. Otospongiosis and sodium fluoride. A clinical double-blind, placebo-controlled study on sodium fluoride treatment in otospongiosis. *Am J Otol* 10:20–22, 1989.
38. Bretlau P, Causse JB, Causse J. Modification with NaF in disequilibrium of otospongiosis origin. *Adv Otorhinolaryngol* 28:73–79, 1982.
39. Bretlau P, Causse J, Causse JB, et al. Otospongiosis and sodium fluoride. A blind experimental and clinical evaluation of the effect of sodium fluoride treatment in patients with otospongiosis. *Ann Otol Rhinol Laryngol* 94:103–107, 1985.
40. Causse J, Causse JB. Clinical studies on fluoride in otospongiosis. *Am J Otol* 6:51–58, 1985.
41. Kerr GS, Hoffman GS. Fluoride therapy for otosclerosis. *Ear Nose Throat J* 68:426–429, 1989.
42. Shea J, Bretlau PM, Hart C, et al. Panel debate for or against fluoride treatment for otosclerosis. *Am J Otol* 6:59–60, 1985.
43. Britton BH. Radiologic aspects in differential diagnosis of neural and sensory deafness. *Ann Otol Rhinol Laryngol* 83:286–293, 1974.
44. Britton BH Jr, Linthicum FH Jr. Otosclerosis: Histologic confirmation of radiologic findings. *Ann Otol Rhinol Laryngol* 79:5–11, 1970.
45. Colletti V, Fiorino FG. Stapedius reflex in the monitoring of NaF treatment of subclinical otosclerosis. *Acta Otolaryngol* 104:447–453, 1987.
46. Lando M, Hoover LA, Finerman G. Stabilization of hearing loss in Paget's disease with calcitonin and etidronate. *Arch Otolaryngol Head Neck Surg* 114:891–894, 1988.
47. Frisch T, Sorensen MS, Overgaard S, Bretlau P. Estimation of volume referent bone turnover in the otic capsule after sequential point labeling. *Ann Otol Rhinol Laryngol* 109:33–39, 2000.
48. Aminpour S, Tinling SP, Brodie HA. Role of tumor necrosis factor-alpha in sensorineural hearing loss after bacterial meningitis. *Otol Neurotol* 26:602–609, 2005.
49. Adams JC. Clinical implications of inflammatory cytokines in the cochlea: A technical note. *Otol Neurotol* 23:316–322, 2002.
50. Fujioka M, Kanzaki S, Okano HJ, et al. Proinflammatory cytokines expression in noise-induced damaged cochlea. *J Neurosci Res* 83:575–583, 2006.
51. Karosi T, Jokay I, Konya J, et al. Detection of osteoprotegerin and TNF-alpha mRNA in ankylotic stapes footplates in connection with measles virus positivity. *Laryngoscope* 116:1427–1433, 2006.
52. Karosi T, Konya J, Szabo LZ, et al. Codetection of measles virus and tumor necrosis factor-alpha mRNA in otosclerotic stapes footplates. *Laryngoscope* 115:1291–1297, 2005.
53. Ren J, Li H, Lu Y. The determinations of tumor necrosis factor and interleukin 6 in serum of patients with sudden sensorineural hearing loss. *J Clin Otorhinolaryngol* 12:311–313, 1998.
54. Satoh H, Firestein GS, Billings PB, et al. Tumor necrosis factor-alpha, an initiator, and etanercept, an inhibitor of cochlear inflammation. *Laryngoscope* 112:1627–1634, 2002.
55. Satoh H, Firestein GS, Billings PB, et al. Proinflammatory cytokine expression in the endolymphatic sac during inner ear inflammation. *J Assoc Res Otolaryngol* 4: 139–147, 2003.
56. Staecker H, Lefebvre PP. Autoimmune sensorineural hearing loss improved by tumor necrosis factor-alpha blockade: a case report. *Acta Otolaryngol* 122:684–687, 2002.
57. Zou J, Pyykko I, Sutinen P, Toppila E. Vibration induced hearing loss in guinea pig cochlea: Expression of TNF-alpha and VEGF. *Hear Res* 202:13–20, 2005.
58. Zehnder AF, Kristiansen AG, Adams JC, et al. Osteoprotegerin knockout mice demonstrate abnormal remodeling of the otic capsule and progressive hearing loss. *Laryngoscope* 116:201–206, 2006.
59. Adachi O, Stankovic KM, Kirstiansen AG, et al. Bisphosphonates inhibit bone remodeling in the otic capsule of osteoprotegerin deficient mouse, an animal model of otosclerosis. Poster presentation 39, meeting of the Association for Research in Otolaryngology, Phoenix, AZ, 2008.
60. Brookes GB. Vitamin D deficiency—a new cause of cochlear deafness. *J Laryngol Otol* 97:405–420, 1983.
61. Brookes GB. Vitamin D deficiency and otosclerosis. *Otolaryngol Head Neck Surg* 93:313–321, 1985.
62. Brookes GB. Vitamin D deficiency and deafness: 1984 update. *Am J Otol* 6:102–107, 1985.
63. Hsu CJ, Lin KN. Activities of carbonic anhydrase in the cochleae of guinea pigs with early experimental endolymphatic hydrops. *Proc Natl Sci Council, Republic of China* 18:107–111, 1994.
64. Drescher DG. Purification of a carbonic anhydrase from the inner ear of the guinea pig. *Proc Natl Acad Sci U S A* 74:892–896, 1977.

65. Lim DJ, Karabinas C, Trune DR. Histochemical localization of carbonic anhydrase in the inner ear. *Am J Otolaryngol* 4:33–42, 1983.
66. Watanabe K, Ogawa A. Carbonic anhydrase activity in stria vascularis and dark cells in vestibular labyrinth. *Ann Otol Rhinol Laryngol* 93:262–266, 1984.
67. Hsu CJ, Nomura Y. Carbonic anhydrase activity in the inner ear. *Acta Otolaryngol Suppl* 418:1–42, 1985.
68. Spicer SS, Schulte BA. Differentiation of inner ear fibrocytes according to their ion transport related activity. *Hear Res* 56:53–64, 1991.
69. Hsu CJ. Ultrastructural study of cytochemical localization of carbonic anhydrase in the inner ear. *Acta Otolaryngol* 111:75–84, 1991.
70. Yamashita H, Sekitani T, Bagger-Sjoberg D. Expression of carbonic anhydrase isoenzyme-like immunoreactivity in the limbus spiralis of the human fetal cochlea. *Hear Res* 64:118–122, 1992.
71. Okamura H, Ohtani I, Sugai N, Suzuki K. The localization and the function of carbonic anhydrase in the inner ear. *Nippon Jibiinkoka Gakkai Kaiho* 96:403–408, 1993.
72. Ichimiya I, Adams JC, Kimura RS. Changes in immunostaining of cochleas with experimentally induced endolymphatic hydrops. *Ann Otol Rhinol Laryngol* 103:457–468, 1994.
73. Ichimiya I, Adams JC, Kimura RS. Immunolocalization of Na⁺, K⁽⁺⁾-ATPase, Ca⁽⁺⁺⁾-ATPase, calcium-binding proteins, and carbonic anhydrase in the guinea pig inner ear. *Acta Otolaryngol* 114:167–176, 1994.
74. Okamura HO, Sugai N, Suzuki K, Ohtani I. Enzyme-histochemical localization of carbonic anhydrase in the inner ear of the guinea pig and several improvements of the technique. *Histochem Cell Biol* 106:425–430, 1996.
75. Gratton MA, Schulte BA, Hazen-Martin DJ. Characterization and development of an inner ear type I fibrocyte cell culture. *Hear Res* 99:71–78, 1996.
76. Spicer SS, Gratton MA, Schulte BA. Expression patterns of ion transport enzymes in spiral ligament fibrocytes change in relation to stria atrophy in the aged gerbil cochlea. *Hear Res* 111:93–102, 1997.
77. Doherty JK, Linthicum FH Jr. Spiral ligament and stria vascularis changes in cochlear otosclerosis: Effect on hearing level. *Otol Neurotol* 25:457–464, 2004.
78. Maier W, Schipper J. Prognostic relevance of anamnestic and diagnostic parameters in low-frequency hearing impairment. *J Laryngol Otol* 120:613–618, 2006.
79. Gates P. Hypothesis: Could Ménière's disease be a chanelopathy? *Intern Med J* 35:488–489, 2005.
80. Vollrath M, Marangos N, Hesse G. Dehydration therapy in low tone hearing loss. An alternative to rheologic therapy? *HNO* 38:154–157, 1990.
81. Ralli G, Celestino D, Fabbriatore M, et al. Effect of acetazolamide on Ménière's disease. *Acta Otorhinolaryngol Ital* 9:503–509, 1989.
82. Blair HC, Schlesinger PH, Ross FP, Teitelbaum SL. Recent advances toward understanding osteoclast physiology. *Clin Orthop* 294:7–22, 1993.
83. Borthwick KJ, Kandemir N, Topaloglu R, et al. A phenocopy of CAII deficiency: A novel genetic explanation for inherited infantile osteopetrosis with distal renal tubular acidosis. *J Med Genet* 40:115–121, 2003.
84. de Vernejoul MC, Benichou O. Human osteopetrosis and other sclerosing disorders: Recent genetic developments. *Calcif Tissue Int* 69:1–6, 2001.
85. Felix R, Hofstetter W, Cecchini MG. Recent developments in the understanding of the pathophysiology of osteopetrosis. *Eur J Endocrinol* 134:143–156, 1996.
86. Jilka RL, Rogers JI, Khalifah RG, Vaananen HK. Carbonic anhydrase isozymes of osteoclasts and erythrocytes of osteopetrotic microphthalmic mice. *Bone* 6:445–449, 1985.
87. Sly WS, Hewett-Emmett D, Whyte MP, et al. Carbonic anhydrase II deficiency identified as the primary defect in the autosomal recessive syndrome of osteopetrosis with renal tubular acidosis and cerebral calcification. *Proc Natl Acad Sci U S A* 80:2752–2756, 1983.
88. Sundquist KT, Vaananen HK, Marks SC Jr. Carbonic anhydrase II and H⁺-ATPase in osteoclasts of four osteopetrotic mutations in the rat. *Histochem Cell Biol* 111:55–60, 1999.
89. Whyte MP. Carbonic anhydrase II deficiency. *Clin Orthop Relat Res* 294:52–63, 1993.
90. Charles DA, Barber HO, Hope-Gill HF. Blood glucose and insulin levels, thyroid function, and serology in Ménière's disease, recurrent vestibulopathy, and psychogenic vertigo. *J Otolaryngol* 8:347–353, 1979.
91. Weille F. Hypoglycemia in Ménière's disease. *Arch Otolaryngol* 84:555–557, 1968.
92. Powers WH. Metabolic aspects of Ménière's disease. *Laryngoscope* 82:1716–1725, 1972.
93. Spencer JT Jr. Hyperlipoproteinemias in the etiology of inner ear disease. *Laryngoscope* 83:639–678, 1973.
94. Spencer JT Jr. Hyperlipoproteinemia and inner ear disease. *W V Med J* 70:215–221, 1974.
95. Spencer JT Jr. Hyperlipoproteinemia and inner ear disease. *Otolaryngol Clin North Am* 8:483–492, 1975.
96. Updegraff W. Impaired carbohydrate metabolism and idiopathic Ménière's disease. *Ear Nose Throat* 56:160–163, 1977.
97. Powers WH. Metabolic aspects of Ménière's disease. *Laryngoscope* 88:122–129, 1978.
98. Kinney SE. The metabolic evaluation in Ménière's disease. *Otolaryngol Head Neck Surg* 88:594–598, 1980.
99. Spencer JT Jr. Hyperlipoproteinemia, hyperinsulinism, and Ménière's disease. *South Med J* 74:1194–1197, 1200, 1981.
100. Kraft JR. Hyperinsulinemia: A merging history with idiopathic tinnitus, vertigo, and hearing loss. *Int Tinnitus J* 4:127–130, 1998.
101. Spencer JT Jr. Blood lipids. *Ear Nose Throat J* 77:224, 1998.
102. Kirtane MV, Medikeri SB, Rao P. Blood levels of glucose and insulin in Ménière's disease. *Acta Otolaryngol* 406:42–45, 1984.

103. Goldman HB. Metabolic causes of fluctuant hearing loss. *Otolaryngol Clin North Am* 8:369–374, 1975.
104. Nowak K, Banaszewski J, Dabrowski P, et al. Tinnitus in systemic diseases. *Otolaryngol Pol* 56:213–216, 2002.
105. Meyerhoff EL. The thyroid and audition. *Laryngoscope* 86:483–489, 1976.
106. Moriyama K, Nozaki M, Kudo J, et al. Sudden deafness in a man with thyrotoxic hypokalemic periodic paralysis. *Jpn J Med* 27:329–332, 1988.
107. Kraft JR. IMx (Abbott) immunoassay of insulin: A practical alternative to RIA hyperinsulinemia identification in idiopathic neurootology and other hyperinsulin metabolic disorders. *Int Tinnitus J* 3:113–116, 1997.
108. Kraft JR. Hyperinsulinemia: The common denominator of subjective idiopathic tinnitus and other idiopathic central and peripheral neurootologic disorders. *Int Tinnitus J* 1:46–53, 1995.
109. Brookler KH. Dizziness, hyperactive caloric responses, otic capsule demineralization, impaired glucose tolerance, and hyperinsulinemia. *Ear Nose Throat J* 85:224, 229, 2006.
110. Brookler KH. Ménière's syndrome, otosclerosis, and insulin resistance syndrome. *Ear Nose Throat J* 85:82–83, 2006.
111. Wrenshall GA, Andrus SB, Mayer J. High levels of pancreatic insulin coexistent with hyperplasia and degranulation of beta cells in mice with the hereditary obese-hyperglycemic syndrome. *Endocrinology* 56:335–340, 1955.
112. Kraft JR. *Diabetes Epidemic and You*. Victoria, BC, Canada: Trafford Publishing, 2008.
113. Frisina ST, Mapes F, Kim S, et al. Characterization of hearing loss in aged type II diabetics. *Hear Res* 211:103–113, 2006.
114. Frittitta L, Youngren J, Vigneri R, et al. PC-1 content in skeletal muscle of non-obese, non-diabetic subjects: Relationship to insulin receptor tyrosine kinase and whole body insulin sensitivity. *Diabetologia* 39:1190–1195, 1996.
115. Johnson KA, Hesse L, Vaingankar S, et al. Osteoblast tissue-nonspecific alkaline phosphatase antagonizes and regulates PC-1. *Am J Physiol Regul Integr Comp Physiol* 279:R1365–1377, 2000.
116. Stefan C, Wera S, Stalmans W, Bollen M. The inhibition of the insulin receptor by the receptor protein PC-1 is not specific and results from the hydrolysis of ATP. *Diabetes* 45:980–983, 1996.
117. Kahn CR, Vicent D, Doria A. Genetics of non-insulin-dependent (type-II) diabetes mellitus. *Annu Rev Med* 47: 509–531, 1996.
118. Frittitta L, Youngren JF, Sbraccia P, et al. Increased adipose tissue PC-1 protein content, but not tumour necrosis factor-alpha gene expression, is associated with a reduction of both whole body insulin sensitivity and insulin receptor tyrosine-kinase activity. *Diabetologia* 40:282–289, 1997.
119. Goldfine ID, Maddux BA, Youngren JF, et al. Membrane glycoprotein PC-1 and insulin resistance. *Mol Cell Biochem* 182:177–184, 1998.
120. Kumakura S, Maddux BA, Sung CK. Overexpression of membrane glycoprotein PC-1 can influence insulin action at a post-receptor site. *J Cell Biochem* 68:366–377, 1998.
121. Goldfine ID, Maddux BA, Youngren JF, et al. Role of PC-1 in the etiology of insulin resistance. *Ann N Y Acad Sci* 892:204–222, 1999.
122. Frittitta L, Camastra S, Baratta R, et al. A soluble PC-1 circulates in human plasma: Relationship with insulin resistance and associated abnormalities. *J Clin Endocrinol Metab* 84:3620–3625, 1999.
123. Groop L. Genetics of the metabolic syndrome. *Br J Nutr* 83(Suppl 1):S39–S48, 2000.
124. Wanic K, Malecki M, Klupa T, et al. TNF-alpha PC-1 gene polymorphisms and pre-diabetes quantitative features in the Polish population. *Przegl Lek* 59:888–891, 2002.
125. Pender C, Ortmeyer HK, Hansen BC, et al. Elevated plasma cell membrane glycoprotein levels and diminished insulin receptor autophosphorylation in obese, insulin-resistant rhesus monkeys. *Metabolism* 51:465–470, 2002.
126. Pender C, Goldfine ID, Kulp JL, et al. Analysis of insulin-stimulated insulin receptor activation and glucose transport in cultured skeletal muscle cells from obese subjects. *Metabolism* 54:598–603, 2005.
127. Goldfine ID, Maddux BA, Youngren JF, et al. The role of membrane glycoprotein plasma cell antigen 1/ectonucleotide pyrophosphatase phosphodiesterase 1 in the pathogenesis of insulin resistance and related abnormalities. *Endocr Rev* 29:62–75, 2008.
128. Ferron M, Hinoi E, Karsenty G, Ducy P. Osteocalcin differentially regulates beta cell and adipocyte gene expression and affects the development of metabolic diseases in wild-type mice. *Proc Natl Acad Sci U S A* 105:5266–5270, 2008.
129. Wolf G. Energy regulation by the skeleton. *Nutr Rev* 66: 229–233, 2008.
130. Strohbach C, Kleinman S, Linkhart T, et al. Potential involvement of the interaction between insulin-like growth factor binding protein (IGFBP)-6 and LIM mineralization protein (LMP)-1 in regulating osteoblast differentiation. *J Cell Biochem* 104:1890–1905, 2008.
131. Li SH, Guo DZ, Li B, et al. The stimulatory effect of insulin-like growth factor-1 on the proliferation, differentiation, and mineralisation of osteoblastic cells from Holstein cattle. *Vet J* 179(3):430–436, 2009.
132. Im JA, Yu BP, Jeon JY, Kim SH. Relationship between osteocalcin and glucose metabolism in postmenopausal women. *Clin Chim Acta Int J Clin Chem* 396:66–69, 2008.
133. Perifanis V, Vyzantiadis T, Tziomalos K, et al. Effect of zoledronic acid on markers of bone turnover and mineral density in osteoporotic patients with beta-thalassaemia. *Ann Hematol* 86:23–30, 2007.
134. Gad HI. The potential osteogenic effects of systemic leptin and insulin administration in streptozotocin-induced diabetic female rats. *Saudi Med J* 28:1185–1190, 2007.

135. Lee NK, Sowa H, Hinoi E, et al. Endocrine regulation of energy metabolism by the skeleton. *Cell* 130:456–469, 2007.
136. Botolin S, McCabe LR. Chronic hyperglycemia modulates osteoblast gene expression through osmotic and non-osmotic pathways. *J Cell Biochem* 99:411–424, 2006.
137. Ix JH, Wassel CL, Kanaya AM, et al. Fetuin-A and incident diabetes mellitus in older persons. *JAMA* 300:182–188, 2008.
138. Stefan N, Fritsche A, Weikert C, et al. Plasma fetuin-A levels and the risk of type 2 diabetes. *Diabetes* 57(10):2762–2767, 2008.
139. Mehrotra R. Emerging role for fetuin-A as contributor to morbidity and mortality in chronic kidney disease. *Kidney Int* 72:137–140, 2007.
140. Mehrotra R. Disordered mineral metabolism and vascular calcification in nondialyzed chronic kidney disease patients. *J Ren Nutr* 16:100–118, 2006.
141. Shroff RC, Shah V, Hiorns MP, et al. The circulating calcification inhibitors, fetuin-A and osteoprotegerin, but not matrix Gla protein, are associated with vascular stiffness and calcification in children on dialysis. *Nephrol Dial Transplant* 23(10):3263–3271, 2008.
142. Messa P, Alberti L, Como G, et al. Calcimimetic increases osteoprotegerin and decreases fetuin-A levels in dialysis patients. *Nephrol Dial Transplant* 22:2724–2725, 2007.
143. Kapadia R, Yi JH, Vemuganti R. Mechanisms of anti-inflammatory and neuroprotective actions of PPAR-gamma agonists. *Front Biosci* 13:1813–1826, 2008.
144. Ichimiya I, Yoshida K, Hirano T, et al. Significance of spiral ligament fibrocytes with cochlear inflammation. *Int J Pediatr Otorhinolaryngol* 56:45–51, 2000.
145. Street I, Jobanputra P, Proops DW. Etanercept, a tumour necrosis factor alpha receptor antagonist, and methotrexate in acute sensorineural hearing loss. *J Laryngol Otol* 120:1064–1066, 2006.
146. Van Wijk F, Staecker H, Keithley E, Lefebvre PP. Local perfusion of the tumor necrosis factor alpha blocker infliximab to the inner ear improves autoimmune neurosensory hearing loss. *Audiol Neurootol* 11:357–365, 2006.
147. Brookler KH. Can a disorder of the vestibular system underlie an etiology for migraine? *Ear Nose Throat J* 87:258, 260–251, 2008.
148. Karosi T, Jokay I, Konya J, et al. Expression of measles virus receptors in otosclerotic, non-otosclerotic and in normal stapes footplates. *Eur Arch Otorhinolaryngol* 264(6):607–613, 2007.
149. Karosi T, Konya J, Szabo LZ, Sziklai I. Measles virus prevalence in otosclerotic foci. *Adv Otorhinolaryngol* 65:93–106, 2007.